

Carbapenemase among Clinical Bacterial Isolates in Nepal

Surya Prasad Devkota,^{1,2} Ashmita Paudel,³ Dharm Raj Bhatta,⁴ Krishna Gurung⁵

¹Pokhara Bigyan Tatha Prabidhi Campus, Nayabazar, Pokhara, ²School of Health and Allied Sciences, Pokhara University, Pokhara, ³Regional College of Health Science and Technology, Nayabazar, Pokhara, ⁴Manipal College of Medical Sciences, Pokhara, ⁵Prithvi Narayan Campus, Pokhara, Nepal.

ABSTRACT

Gram-negative bacterial isolates producing carbapenemase enzymes are great public health problem in developing countries and their control is challenging task due to the involvement of multiple factors including the practice of self-medication, use of antibiotics on animal farms, poor hospital hygiene, etc. In this review, we searched various relevant publications on carbapenemase-producing bacterial isolates in Nepal.

Various classes of carbapenemase producing bacteria have been reported in Nepal. Most frequent was the New Delhi Metallo beta lactamase with many variants. Similarly, oxacillinase and *Klebsiella pneumoniae* carbapenemase producers were also prevalent in Nepal. Likewise, other carbapenemase like Verona integron-encoded metallo- β -lactamase, imipenemase, and Dutch imipenemase were also detected. Ten variants of class B carbapenemase were detected including seven variants of New Delhi Metallo- β -lactamase and one variant each of Verona integron-encoded metallo- β -lactamase, Dutch imipenemase, and imipenemase. Similarly, 12 variants of oxacillinase were reported while no variants of *Klebsiella pneumoniae* carbapenemase were reported. The isolates producing carbapenemase were extremely drug-resistant as they also co-produced various other carbapenemase, beta-lactamase, 16S rRNA methylase. Such isolates had very few treatment options as only last line drugs like colistin, fosfomycin, and tigecycline were effective against most of these isolates. Carbapenemase production by almost all gram-negative pathogens is a matter of concern because some of these enzymes are located on plasmids and pose rapid dissemination among various gram-negative pathogens. Timely surveillance for carbapenemase producers throughout the nation, their proper treatment, and proper hospital hygiene to prevent nosocomial infections by carbapenemase producers, controlled use of carbapenems, educating health care workers, students, and the general public about the adverse effects of antimicrobial resistance are imminent.

Keywords: Carbapenemase; KPC; NDM; Nepal; OXA.

INTRODUCTION

Carbapenemase are the member of β -lactamase having the flexible ability of β -lactam hydrolysis.¹ Carbapenemase are the global public health challenge since their discovery and are classified as class A, B, and D carbapenemase as per Ambler classification.² Class A carbapenemases are serine carbapenemases as they must contain serine for their hydrolytic activity. *Klebsiella pneumoniae* carbapenemase (KPC), *Serratia marcescens* enzyme (SME), Guiana extended spectrum β -lactamase (GES), and imipenem-hydrolyzing β -lactamase (IMI) are the major genes in this class of carbapenemases with many variants.¹ Metallo- β -lactamases or class B carbapenemases contain metal ions at the active site

of enzymes. New Delhi Metallo- β -lactamase (NDM), Verona integron-encoded metallo- β -lactamase (VIM), and imipenemase (IMP) are the significant genes of this carbapenemase.² Class D carbapenemases or oxacillin-hydrolyzing β -lactamases (OXA) carbapenemases are common in *Acinetobacter baumannii*. OXA-23, OXA-24, OXA-51, OXA-55, OXA-58, OXA-48, OXA-50, OXA-60, and OXA-62 are the major subgroups.¹

Carbapenem-resistant isolates also carry various other resistance determinants in many instances limiting the options for their treatment. Not only this, the mortality rate is higher in case of infections by these pathogens in comparison to carbapenem susceptible isolates.³ Excess use of carbapenem for the treatment of ESBL

Correspondence: Surya Prasad Devkota, Pokhara Bigyan Tatha Prabidhi Campus, Nayabazar, Pokhara. Email: devkotasp1@gmail.com, Phone: +9779846434924.

producers is the cause of carbapenem resistance and this resistance mechanism is more severe in *Klebsiella* with more than 50 % prevalence in South East Asia.⁴

Prevalence of carbapenemase-producing gram-negative isolates is high in Nepal but there are very few studies about these pathogens. As a result various characteristics of these isolates are less known in our country. Hence, this study was carried out to sum up the information about these isolates.

METHODS

A systematic literature search was done for various carbapenemase including class B (NDM, VIM, IMP, SPM, GIM, SIM), class A (KPC, SME, IMI, GES, NMC), and class D (OXA) carbapenemase genes and enzymes in Nepal from various electronic databases (Medline via PubMed, Embase, NepJOL and other databases) published till January 2019. Original research articles, as well as review articles available in English indicating any of class A, B and D carbapenemase detection using both phenotypic and molecular methods from Nepal, were included in this study (Figure 1). Various keywords used for literature search in the abstract and title of the articles were: NDM gene/enzyme, VIM gene/enzyme, IMP gene/enzyme, SPM gene/enzyme, GIM gene/enzyme, SIM gene/enzyme, KPC gene/enzyme, SME gene/enzyme, IMI gene/enzyme, GES gene/enzyme, NMC gene/enzyme, OXA gene/enzyme, Nepal, class A carbapenemase, class B carbapenemase, class D carbapenemase, multi-drug resistant, gram-negative isolates, prevalence.

Research articles containing following information in text and/ or abstract were selected in the study; i) reported various carbapenemase producing gram-negative bacterial pathogens that were isolated in Nepal, ii) used standard phenotypic and molecular techniques for the detection of various carbapenemase producers, iii) included various properties of carbapenemase producers like duration of study, isolate producing carbapenemase, variants of the carbapenemases, site of study, prevalence and antibiotic susceptibility profile. Similarly, the exclusion criteria were; i) not used the standard phenotypic (carbapenemase inhibition methods using EDTA, boronic acid derivatives, and dipicolinic acid) and molecular method (PCR and/ gene sequencing) for carbapenemase detection, ii) not included various characteristics of carbapenemase producers, iii) duplicate articles of one study, iv) meta-analysis, v) articles on languages other than English, and vi) articles containing abstract only.

DATA EXTRACTION

Variables extracted from the selected studies were:

carbapenemase positive isolate, source specimen, study period, co-existence of other resistant determinants, variants of class A, B and D carbapenemase, the prevalence of carbapenemase producers, study site and antibiotic susceptibility profile. Selected articles were independently reviewed by four reviewers fairly and consensus was made to solve any inconsistencies among reviewers.

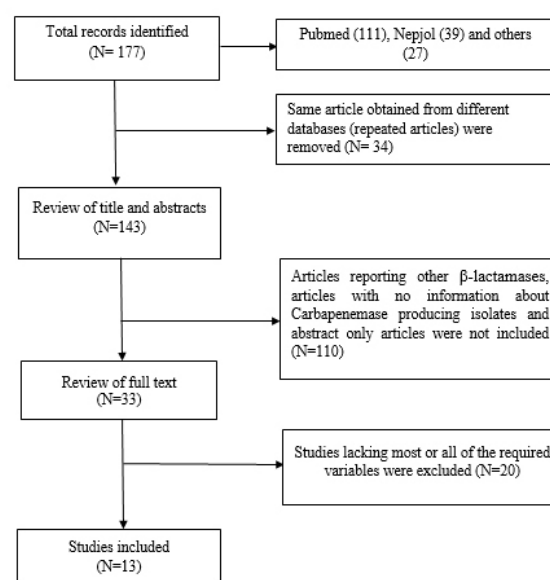


Figure 1. selection process of the included studies.

RESULTS

Various carbapenemase producing Gram-negative pathogens have been reported in Nepal including class A, B, and D carbapenemase using both molecular and phenotypic methods. Major carbapenemase reported were NDM, OXA, KPC, VIM, and IMP. Significant carbapenemase producers reported as per the articles reviewed were *E. coli*, *Klebsiella* spp, *Acinetobacter baumannii*, *Pseudomonas aeruginosa*, and *Providencia rettgeri*. Almost all carbapenemase bearing isolates were positive for many other resistance determinants and all were multidrug resistant in nature.

CLASS A CARBAPENEMASE

Only *Klebsiella pneumoniae* carbapenemase (KPC) had been reported in Nepal among *E. coli*, *K. pneumoniae*, and *A. baumannii* isolates with the prevalence range of 0.8 to 8 percentages. All studies detected this resistance mechanism using a phenotypic method using imipenem and imipenem/phenylboronic acid. Most of these isolates were also co-producing metallo beta-lactamase enzyme increasing their drug resistance pattern. Other subclasses of this carbapenemase like SME, IMI, GES, NMC, etc. have not been detected yet and there is imminent need of screening for these carbapenemases.

Table 1. Characteristics of Class A Carbapenemase producing Gram-negative isolates.

Subclass	Isolates	Study period	Prevalence among total GNB studied (%)	Other resistance genes/factors	Study site*	Ref.
KPC	<i>Enterobacteriaceae</i>	2013-14	1.3 (4/310)	MBL	KCH	5
KPC	<i>E. coli</i> , <i>K. pneumoniae</i> , <i>A. baumannii</i>	2013-14	0.8 (6/769)	MBL	KMH	6
KPC	<i>Klebsiella Acinetobacter</i>	2014-15	4.2 (7/165)	NA	TUTH	7
KPC	<i>Acinetobacter</i>	2018	8 (9/111)	MBL	MTH	8
SME, IMI, GES, NMC	NR	NR	NR	NR	NR	NR

*KCH-Kanti Children Hospital, KMH- Kathmandu Model Hospital, TUTH-Tribhuvan University Teaching Hospital, MTH-Manipal Teaching Hospital, MBL- Metallo beta-lactamase enzyme, NA-Not available, NR-Not reported

CLASS B CARBAPENEMASE

Four subclasses of this carbapenemase have been reported in Nepal including NDM, VIM, IPM, and DIM. NDM was the most common carbapenemase enzyme reported with seven variants. While only two variant of VIM have been reported. The highest prevalence of VIM, NDM, and IMP were 18.3%, 6.5%, and 6.1% respectively. Many human pathogens including *E. coli*, *Klebsiella*,

Acinetobacter, *Pseudomonas*, *Enterobacter*, *Citrobacter*, *Proteus*, and *Providencia* spp. were positive for various class B carbapenemase genes. These isolates were highly drug-resistant as most of these isolates were resistant to imipenem, meropenem, aztreonam, ceftazidime, amikacin, arbekacin, ciprofloxacin while most of them were sensitive only to colistin.^{9,11} Subclasses like SPM, GIM, SIM were not prevalent among gram-negative pathogens of Nepal.

Table 2. Characteristics of Class B Carbapenemase producing Gram-negative isolates.

Sub class	Variants	isolates	Study period	Prevalence (%)	Source specimen	Other resistance genes	Study site*	Ref.
NDM	NA	<i>E. coli</i> , <i>Klebsiella</i> , <i>Acinetobacter</i> , <i>Pseudomonas</i> , <i>Enterobacter</i> , <i>Citrobacter</i> , <i>Proteus</i> , <i>Providencia</i>	2015-16	6.5	Urine, pus, sputum blood	NA	MTH	9
NDM	NDM-1	<i>P. rettgeri</i>	2012	NA	Pus Sputum	<i>bla</i> _{OXA-10} , <i>aadA1</i> , <i>bla</i> _{VEB-1} , <i>bla</i> _{TEM} , <i>bla</i> _{ADC-67} , <i>armA</i>	TUTH	10
NDM and VIM	NDM-1 VIM-2	<i>P. aeruginosa</i>	2012-13	NA		RmtB4, PDC-35	TUTH	11
NDM IPM, VIM	NDM-1	<i>E. coli</i>	2012-13	2.15, 2.15, 3.22	NA	<i>bla</i> _{TEM} , <i>bla</i> _{SHV} , <i>bla</i> _{CTX-M}	KMC	12
NDM	NDM-1, NDM-3, NDM-4, NDM-7, NDM-8, NDM-12, and NDM-13	<i>E. coli</i> , <i>Klebsiella</i> , <i>Acinetobacter</i>	2012-15	NA	Urine, pus, sputum blood	<i>bla</i> OXA-23, <i>bla</i> OXA-69, <i>aacC1</i> , <i>aadA1</i> , <i>aadA5armA</i> , <i>bla</i> OXA-32, <i>bla</i> OXA-420, <i>bla</i> PSE-1, <i>aacA2bla</i> OXA-104, <i>bla</i> PER-7, <i>RmtC</i> , <i>bla</i> OXA-94, <i>bla</i> CTX-M-15, <i>bla</i> OXA-181, <i>bla</i> TEM-166, <i>rmtB</i> , <i>aadA2</i> , <i>RmtF</i> , <i>aphA6</i> , <i>OXA-72</i> , <i>SHV-158</i> , <i>TEM-1</i> , <i>SHV-28</i> , <i>SHV-11</i> , <i>SHV-1</i> ,	TUTH PH	13
DIM	DIM-1	<i>P. aeruginosa</i>	2012-13	NA	NA	PDC-32, RmtF2	TUTH	11
VIM	VIM-2	<i>P. aeruginosa</i>	2015-16	18.3	Pus, Urine, Sputum		ANIAS	14
IMP	IMP-1	<i>P. aeruginosa</i>	2015-16	6.1	Pus Urine Sputum		ANIAS	14
SPM, GIM, SIM	NR	NR	NR	NR	NR		NR	NR

CLASS D CARBAPENEMASE

Twelve variants of OXA genes have been reported among *E. coli*, *Providencia rettgeri*, *A. baumannii*, and *K. pneumoniae*. These isolates also possessed various other

resistance factors like beta-lactamase, carbapenemase and methyl transferase. *P. rettgeri*, an OXA-48 positive isolate from Tribhuvan University Teaching Hospital was resistant to all drugs tested (pan-drug resistant).

Table 3. Characteristics of Class D Carbapenemase producing Gram-negative isolates.

Subclass	Variants	Isolates	Study period	Prevalence	Source specimen	Other resistance genes	Study site*	Ref.
OXA	OXA-181	<i>E. coli</i>	2013-14	0.4	NA	NDM-5, CTX-M-15	TUTH	15
OXA	OXA-23 OXA-58	<i>A. baumannii</i>	2013-14	NA	NA	NDM-1, <i>armA</i>	TUTH	16
OXA	OXA-48	<i>Providencia rettgeri</i>	2015	NA	Urine	NA	TUTH	17
OXA	OXA-23, OXA-51 OXA-10 OXA-72 OXA-69 OXA-32 OXA-420 OXA-104 OXA-94	<i>Providencia rettgeri</i> , <i>Acinetobacter K. pneumoniae</i>	2012-15	NA	Pus, sputum, urine	<i>aacC1</i> , <i>aadA1</i> , <i>aadA5</i> , <i>armA</i> , <i>blaPSE-1</i> , <i>aacA2</i> , <i>blaPER-7</i> , <i>RmtC</i> , <i>blaCTX-M-15</i> , <i>blaTEM-166</i> , <i>rmtB</i> , <i>aadA2</i> , <i>RmtF</i> , <i>aphA6</i> , <i>SHV-158</i> , <i>TEM-1</i> , <i>SHV-28</i> , <i>SHV-11</i> , <i>SHV-1</i> , <i>NDM-1</i> , <i>NDM-3</i> , <i>NDM-4</i> , <i>NDM-7</i> , <i>NDM-8</i> , <i>NDM-12</i> , and <i>NDM-13</i>	TUTH	13

DISCUSSION

Based on the articles reviewed, all three types of carbapenemase-producing pathogens have been detected in Nepal since the beginning of this decade. Carbapenemase positive isolates were reported from various specimens including urine, pus, sputum, and blood. Almost all significant gram-negative pathogens were capable of producing these resistance factors. Most common isolates producing carbapenemase were *E. coli*, *K. pneumoniae*, *A. baumannii*, *P. aeruginosa*, *P. rettgeri* with a prevalence range of 0.4 to 18.3%. Most of the carbapenem producing isolates also coproduced a wide variety of other drug-resistance genes including metallo beta-lactamases, extended spectrum beta-lactamase, acetyltransferase, methyltransferase, aminoglycoside phosphotransferase, streptomycin adenylyltransferase, beta-lactamase, etc. Due to the coproduction of these resistance determinants almost all carbapenemase bearing isolates were highly drug-resistant.

Among various subclasses of class A carbapenemase, only the KPC has been reported. KPC was not limited only in *K. pneumoniae* but also reported in *E. coli* and *A. baumannii*. Other studies also reported KPC from different pathogens including *Citrobacter*, *Salmonella*, *Enterobacter*, *E. coli*, *Proteus*, *Serratia*, *Pseudomonas*, *Acinetobacter*, and *K. oxytoca*,¹⁸ *Morganella morganii*¹⁹,

Providencia stuartii,²⁰ and *Enterococcus spp.* and *Shigella spp.*²¹ The data also showed that the prevalence of KPC among gram-negative pathogens is increasing continuously as the incidence was 0.8% in 2013 while it was 8% in 2018. This increase of KPC producers is a matter of concern as there is no regular screening of such pathogens in clinical settings of Nepal and may cause epidemic outbreaks if not monitored properly. These isolates also co-produced metallo beta-lactamase enzymes leading to increased drug resistance. Co-production of other drug resistance genes is common among these isolates.¹⁸ Likewise, KPC producers were found to co-harbor *bla*_{CTX-M-15}, *bla*_{TEM-1} and *bla*_{SHV-11},²² *NDM-1*²³, *mcr-1*^{24,25}, and *VIM*^{26,27}. Most studies of these isolates were focused only at Kathmandu valley and very less in other parts of the nation hence there is imminent need of their study throughout the country. Similarly, study of subclasses like SME, IMI, GES, NMC is not done in Nepal and this may lead to unnoticed dissemination of pathogens producing such genes.

NDM, VIM, IPM, and DIM were the reported subclasses of Class B Carbapenemase in Nepal. Among them, NDM was the most predominant one with respect to both cases reported and the number of variants. Almost all major gram-negative pathogens were positive for NDM gene which may facilitate its spread to other human pathogens as most of the isolates bear NDM gene in plasmids. Wide dissemination of NDM gene among

variety of human pathogens via plasmids is also reported earlier.^{28,29} VIM, IPM, and DIM genes were also detected in Nepal but less frequently in comparison of NDM. Most of the class B carbapenemase producers also produced a wide variety of other resistance determinants conferring resistance to nearly all commonly used antibiotics. Many other antibiotic resistant factors co-exist with class B carbapenemase bearing isolates to make them resistant to wider range of antibiotics³⁰⁻³⁷ and some of these isolates were also pan-drug resistant.³⁰ Only very few treatment options were there for these isolates. Extreme drug resistance among these isolates with very limited treatment options has been reported already.³⁸⁻⁴⁰ Tigecycline and colistin,⁴¹ fosfomycin⁴² are the available treatment options for class B and other carbapenemases. However, there was the report of colistin and fosfomycin resistance among NDM bearing *P. rettgeri* isolated in Kathmandu in 2012.¹⁰ Extreme drug resistance of these isolates is a serious problem for their proper treatment. In addition to this these isolates have been associated with nosocomial outbreaks further complexing their proper management.⁴³⁻⁴⁶

Many variants of oxacillinases have been reported after 2013. This carbapenemase is not limited only in *Acinetobacter* as there are the reports of oxacillinase detection among *Providencia rettgeri*, *K. pneumoniae*, and *E.coli*. This fact indicates that these pathogens are widespread among many members of Enterobacteriaceae in our nation. These isolates are widespread in *P. aeruginosa* and *A. baumannii* while less frequent in members of Enterobacteriaceae.⁴⁷ They have also been reported from *Acinetobacter*, *P. aeruginosa*, *Shewanella*, *Ralstonia pickettii*, *Pandoraea pnomenus* and members of Enterobacteriaceae.⁴⁸ OXA positive isolates were also found to be carrying many other genes conferring resistance to other drugs including NDM, CTX-M-15, armA and others. Various carbapenemase genes frequently associated with OXA genes are NDM⁴⁹, NDM-1 and KPC.⁵⁰ Pan-drug resistance was also reported on one isolate carrying OXA-48 indicating no treatment option for such isolates. According to the study of Tavares et al the majority of OXA positive isolates were extremely drug-resistant indicating their highly elevated drug resistance.⁵¹

CONCLUSIONS

All three classes of carbapenemase have been detected in Nepal and those isolates were highly drug-resistant. Timely surveillance of these pathogens in clinical settings, detail molecular analysis and proper use of antibiotics is urgent.

REFERENCES

1. Queenan AM, Bush K. Carbapenemases: the versatile β -lactamases. Clin Microbiol Rev. 2007;20(3):440-58. [\[Link\]](#)
2. Bedenić B, Plečko V, Sardelić S, Uzunović S, Torkar KG. Carbapenemases in gram-negative bacteria: Laboratory detection and clinical significance. Biomed Res Int. 2014;2014:841951. [\[Link\]](#)
3. Lutgring JD, Limbago BM. The problem of carbapenemase-producing-carbapenem-resistant-Enterobacteriaceae detection. J Clin Microbiol. 2016;54(3):529-534. [\[Link\]](#)
4. Teo JQM, Cai Y, Lim TP, Tan TT, Kwa ALH. Carbapenem resistance in gram-negative bacteria: The not-so-little problem in the little red dot. Microorganisms. 2016;4(1):13. [\[PubMed\]](#)
5. Gautam K, Lekhak B, Triphatee PP, Bhatta DR. Phenotypic detection of carbapenemases and β -lactamases induced carbapenem resistance in Enterobacteriaceae. The Pharmaceutical and Chemical Journal. 2015;2(2):1-7. [\[FullText\]](#)
6. Karn S, Pant ND, Neupane S, Khatiwada S, Basnyat S, Shrestha B. Prevalence of carbapenem resistant bacterial strains isolated from different clinical samples: study from a tertiary care hospital in Kathmandu, Nepal. Journal of Biomedical Sciences. 2016;3(1):11-5. [\[FullText\]](#)
7. Parajuli NP, Acharya SP, Mishra SK, Parajuli K, Rijal BP, Pokhrel BM. High burden of antimicrobial resistance among gram negative bacteria causing healthcare associated infections in a critical care unit of Nepal. Antimicrob Resist Infect Control. 2017;6:67. [\[FullText\]](#)
8. Bhatta DR, Hamal D, Shrestha R, Supram HS, Joshi P, Nayak N, et al. Burden of multidrug resistant respiratory pathogens in intensive care units of tertiary care hospital. Asian Journal of Medical Sciences. 2019;10(2):14-19. [\[FullText\]](#)
9. Devkota SP, Sharma S, Bhatta DR, Paudel A, Sah AK, Kandel BP. Prevalence of the blaNDM gene among metallo- β -lactamase-producing Gram-negative isolates from western Nepal. J Glob Antimicrob Resist. 2018;12:3-4. [\[DOI\]](#) [\[ScienceDirect\]](#)
10. Tada T, Miyoshi-Akiyama T, Dahal RK, Sah MK, Ohara H, Shimada K et al. NDM-1 Metallo- β -Lactamase and ArmA 16S rRNA methylase producing *Providencia rettgeri* clinical isolates in Nepal. BMC Infect Dis. 2014;14:56. [\[PubMed\]](#)
11. Tada T, Shimada K, Satou K, Hirano T, Pokhrel BM, Sherchand JB, et al. *Pseudomonas aeruginosa* clinical isolates in Nepal coproducing metallo- β -lactamases

- and 16SrRNA methyltransferases. Antimicrob Agents Chemother. 2017;61(9):e00694-17. [\[Link\]](#)
12. Pokhrel RH, Thapa B, Kafle R, Shah PK, Tribuddharat C. Co-existence of beta-lactamases in clinical isolates of *Escherichia coli* from Kathmandu, Nepal. BMC Res Notes. 2014;7:694. [\[Link\]](#)
 13. Devkota SP, Paudel A. New Delhi metallo- β -lactamase among clinical isolates in Nepal. Etorium J Microbiol. 2018;4:100010M08SD2018. [\[FullText\]](#)
 14. Acharya M, Joshi PR, Thapa K, Aryal R, Kakshapati T, Sharma S. Detection of metallo- β -lactamases-encoding genes among clinical isolates of *Pseudomonas aeruginosa* in a tertiary care hospital, Kathmandu, Nepal. BMC Res Notes. 2017;10:718. [\[Link\]](#)
 15. Basudha S, Tada T, Shimada K, Shrestha S, Ohara H, Pokhrel BM, et al. Emergence of various NDM-type-metallo- β -lactamase-producing *Escherichia coli* clinical isolates in Nepal. Antimicrob Agents Chemother. 2017;61:e01425-17. [\[Link\]](#)
 16. Shrestha S, Tada T, Miyoshi-Akiyama T, Ohara H, Shimada K, Satou K, et al. Molecular epidemiology of multidrug-resistant *Acinetobacter baumannii* isolates in a university hospital in Nepal reveals the emergence of a novel epidemic clonal lineage. Int J Antimicrob Agents. 2015;46(5):526-31. [\[ScienceDirect\]](#)
 17. Sah R, Khadka S, Pokharel BM, Sherchand JB, Parajuli K, Shah NP, et al. Detection of OXA-48 (Oxacillinase) producing *Providencia* in an ICU patient for the first time in Nepal. Second National Summit of Health and Population Scientists in Nepal 2016. [\[FullText\]](#)
 18. Arnold RS, Thom KA, Sharma S, Phillips M, Johnson JK, Morgan DJ. Emergence of *Klebsiella pneumoniae* carbapenemase (KPC)-producing bacteria. South Med J. 2011;104:40-5. [\[PubMed\]](#)
 19. Cai JC, Yang W, Hu YY, Zhang R, Zhou HW, Chen GX. Detection of KPC-2 and qnrS1 in clinical isolates of *Morganella morganii* from China. Diagn Microbiol Infect Dis. 2012;73(2):207-9. [\[PubMed\]](#)
 20. Aires CA, Almeida AC, Vilela MA, Morais-Junior MA, Morais MM. Selection of KPC-2-producing *Providencia stuartii* during treatment for septicemia. Diagn Microbiol Infect Dis. 2016;84(1):95-96. [\[PubMed\]](#)
 21. Yang F, Mao D, Zhou H, Luo Y. Prevalence and fate of carbapenemase genes in a wastewater treatment plant in Northern China. PLoS ONE. 2016; 11(5):e0156383. [\[PlosOne\]](#)
 22. Rhee JY, Park YK, Shin JY, Choi JY, Lee MY, Peck KR, et al. KPC-producing extreme drug-resistant *Klebsiella pneumoniae* isolate from a patient with diabetes mellitus and chronic renal failure on hemodialysis in South Korea. Antimicrob Agents Chemother. 2010;54(5):2278-9. [\[Link\]](#)
 23. Wu W, Espedido B, Feng Y, Zong Z. *Citrobacter freundii* carrying bla_{KPC-2} and bla_{NDM-1} : characterization by whole genome sequencing. Sci Rep. 2016;6:30670. [\[Link\]](#)
 24. Mendes AC, Novais Á, Campos J, Rodrigues C, Santos C, Antunes P, et al. *Mcr-1* in carbapenemase-producing *Klebsiella pneumoniae* with hospitalized patients, Portugal, 2016-2017. Emerg Infect Dis. 2018;24:762-6. [\[PubMed\]](#)
 25. Tacão M, Tavares RDS, Teixeira P, Roxo I, Ramalheira E, Ferreira S, et al. *mcr-1* and bla_{KPC-3} in *Escherichia coli* sequence type 744 after meropenem and colistin therapy, Portugal. Emerg Infect Dis. 2017;23:1419-21. [\[PubMed\]](#)
 26. van Dijk K, Voets GM, Scharringa J, Voskuil S, Fluit AC, Rottier WC, et al. A disc diffusion assay for detection of class A, B and OXA-48 carbapenemases in Enterobacteriaceae using phenyl boronic acid, dipicolinic acid and temocillin. Clin Microbiol Infect. 2014;20:345-9. [\[FullText\]](#)
 27. Piedra-Carrasco N, Fabrega A, Calero-Caceres W, Cornejo-Sanchez T, Brown-Jaque M, Mir-Cros A, et al. Carbapenemase-ing *Enterobacteriaceae* recovered from a Spanish river ecosystem. PLoS One. 2017;12:e0175246. [\[PubMed\]](#)
 28. Li X, Fu Y, Shen M, Huang D, Du X, Hu Q, et al. Dissemination of bla_{NDM-5} gene via an IncX3-type plasmid among non-clonal *Escherichia coli* in China. Antimicrob Resist Infect Control. 2018;7:59. [\[PubMed\]](#)
 29. Nithya N, Remitha R, Jayasree PR, Faisal M, Manish Kumar PR. Analysis of beta-lactamases, bla_{NDM-1} phylogeny & plasmid replicons in multidrug-resistant *Klebsiella* spp. from a tertiary care centre in south India. Indian J Med Res. 2017;146:S38-S45. [\[PubMed\]](#)
 30. Chen Z, Qiu S, Wang Y, Wang Y, Liu S, Wang Z, et al. Coexistence of bla_{NDM-1} with the prevalent bla_{OXA23} and bla_{IMP} in pan-drug resistant *Acinetobacter baumannii* Isolates in China. Clin Infect Dis. 2011; 52; 692-693. [\[PubMed\]](#)
 31. Hu L, Zhong Q, Shang Y, Wang H, Ning C, Li Y, et al. The prevalence of carbapenemase genes and plasmid-mediated quinolone resistance determinants in carbapenem-resistant Enterobacteriaceae from five teaching hospitals in central China. Epidemiol Infect. 2014;142(9):1972-7. [\[PubMed\]](#)
 32. Yucel FY, Cakirlar FK, Koyuncu E, Ozturk R. Detection of IMP, VIM and NDM metallo-beta-lactamase carbapenemase genes in carbapenem resistant *Pseudomonas* strains from bloodstream infections in Istanbul, Turkey. Int J Infect Dis. 2016;45:99. [\[FullText\]](#)
 33. Zhong LL, Zhang YF, Doi Y, Huang X, Zhang XF, Zeng KJ, et al. Coproduction of MCR-1 and NDM-1 by colistin-

- resistant *Escherichia coli* isolated from a healthy individual. *Antimicrob Agents Chemother.* 2017;61:e01962-16.[\[Link\]](#)
34. Zowawi HM, Sartor AL, Balkhy HH, Walsh TR, Al Johani SM, Al Jindan RY, et al. Molecular characterization of carbapenemase-producing *Escherichia coli* and *Klebsiella pneumoniae* in the countries of the Gulf Cooperation Council: Dominance of OXA-48 and NDM producers. *Antimicrob Agents Chemother.* 2014;58(6):3085-3090.[\[Link\]](#)
 35. Moubareck CA, Mouftah SF, Pál T, Ghazawi A, Halat DH, et al. Clonal emergence of *Klebsiella pneumoniae* ST14 co-producing OXA-48-type and NDM carbapenemases with high rate of colistin resistance in Dubai, United Arab Emirates. *Int J Antimicrob Agents.* 2018;52:90–95.[\[PubMed\]](#)
 36. Leski TA, Bangura U, Jimmy DH, Ansumana R, Lizewski SE, Li RW, et al. Identification of *bla*_{OXA-51-like}, *bla*_{OXA-58}, *bla*_{DIM-1}, and *bla*_{VIM} carbapenemase genes in hospital *Enterobacteriaceae* isolates from Sierra Leone. *J Clin Microbiol.* 2013;51:2435-2438.[\[PubMed\]](#)
 37. Arana DM, Ortega A, González-Barberá E, Lara N, Bautista V, Gómez-Ruiz D, et al. Carbapenem-resistant *Citrobacter* spp. isolated in Spain from 2013 to 2015 produced a variety of carbapenemases including VIM-1, OXA-48, KPC-2, NDM-1 and VIM-2. *J Antimicrob Chemother.* 2017;72:3283-7.[\[Link\]](#)
 38. Wei WJ, Yang HF, Ye Y, Li JB. New Delhi metallo- β -lactamase-mediated carbapenem resistance: origin, diagnosis, treatment and public health concern. *Chin Med J.* 2015;128:1969-76.[\[PubMed\]](#)
 39. Kus JV, Tadros M, Simor A, Low DE, McGeer AJ, Willey BM, et al. New Delhi metallo- β -lactamase-1: local acquisition in Ontario, Canada, and challenges in detection. *CMAJ.* 2011;183(11):1257-61.[\[PubMed\]](#)
 40. Qin S, Qi H, Zhang Q, Zhao D, Liu ZZ, Tian H, et al. Emergence of extensively drug-resistant *Proteus mirabilis* harboring a conjugative NDM-1 plasmid and a novel *Salmonella* genomic island 1 variant, SGI1-Z. *Antimicrob Agents Chemother.* 2015;59:6601-4.[\[PubMed\]](#)
 41. Shakil S, Ahmad A, Tabrez S, Ashraf GM, Khan AAP. A region specific treatment strategy to address the problem of drug resistance by NDM-1-producing pathogens. *Enz Eng.* 2013;2:107.
 42. Tzouveleki LS, Markogiannakis A, Piperaki E, Souli M, Daikos GL. Treating infections caused by carbapenemase-producing *Enterobacteriaceae*. *Clin Microbiol Infect.* 2014;20(9):862–72.[\[FullText\]](#)
 43. de Jager P, Chirwa T, Naidoo S, Perovic O, Thomas J. Nosocomial outbreak of New Delhi metallo- β -lactamase-1-producing gram-negative bacteria in South Africa: a case-control study. *PLoS ONE.* 2015;10(4):e0123337.[\[FullText\]](#)
 44. Pagani L, Colino C, Migliavacca R, Labonia M, Docquier JD, Nucleo E, et al. Nosocomial outbreak caused by multidrug-resistant *Pseudomonas aeruginosa* producing IMP-13 metallo-beta-Lactamase. *J Clin Microbiol.* 2005; 43:3824-8.[\[FullText\]](#)
 45. Nastro M, Monge R, Zintgraff J, Vaulet LG, Boutourel M, Famiglietti A, et al. First nosocomial outbreak of VIM-16-producing *Serratia marcescens* in Argentina. *Clin Microbiol Infect.* 2012;19(7):617-9.[\[FullText\]](#)
 46. Amoureux L, Riedweg K, Chapuis A, Bador J, Siebor E, Péchinot A, et al. Nosocomial infections with IMP-19-producing *Pseudomonas aeruginosa* linked to contaminated sinks, France. *Emerg Infect Dis.* 2017;23:304-307.[\[PubMed\]](#)
 47. Bialvaei AZ, Kafil HS, Leylabadlo HE, Asgharzadeh M, Aghazadeh M. Dissemination of carbapenemases producing gram negative bacteria in the Middle East. *Iran J Microbiol.* 2015; 7: 226–246.[\[PubMed\]](#)
 48. Sahuquillo-Arce JM, Hernández-Cabezas A, Yarad-Awad F, Ibáñez-Martínez E, Falomir-Salcedo P, Ruiz-Gaitán A. Carbapenemases: a worldwide threat to antimicrobial therapy. *World J Pharmacol.* 2015;4(1):75–95.[\[Link\]](#)
 49. Regeen H, Al-Sharafa-Kittaneh D, Kattan R, Al-Dawodi R, Marzouqa H, Hindiyeh MY. First Report of *bla*_{NDM} and *bla*_{OXA-58} coexistence in *Acinetobacter junii*. *J Clin Microbiol.* 2014;52(9):3492-3.[\[Link\]](#)
 50. Begum N, Shamsuzzaman SM. Emergence of carbapenemase-producing urinary isolates at a tertiary care hospital in Dhaka, Bangladesh. *Ci Ji Yi Xue Za Zhi.* 2016; 28(3): 94-8.[\[PubMed\]](#)
 51. Tavares LCB, Vasconcelos FM, Sousa WV, Rocchetti TT, Mondelli AL, Ferreira AM, et al. Emergence and persistence of high-risk clones among MDR and XDR *A. baumannii* at a Brazilian teaching hospital. *Front Microbiol.* 2019;9:2898.[\[FullText\]](#)